

UltraClean[®] Microbial DNA Isolation Kit

| Catalog No. | Quantity |
|-------------|-----------|
| 12224-50 | 50 Preps |
| 12224-250 | 250 Preps |

Instruction Manual

New protocol instruction: Shake Solution MD3 to mix before using to ensure consistent results.





Table of Contents

| Introduction | 3 |
|--|----|
| Protocol Overview | 3 |
| Flow Chart | 4 |
| Equipment Required | 5 |
| Kit Contents & Storage | 5 |
| Precautions & Warnings | 5 |
| Protocols: | |
| Experienced User Protocol | 6 |
| Detailed Protocol (Describes what is happening at each step) | 7 |
| Vacuum Manifold Protocol1 | 10 |
| Hints & Troubleshooting Guide | 12 |
| Contact Information | 13 |
| Other Quality Products Available | 14 |



Introduction

The UltraClean[®] Microbial DNA Isolation Kit is designed to isolate high-quality genomic DNA from microorganisms. A variety of microorganisms, including bacterial and fungal spores, have been tested successfully with this kit.

Protocol Overview

Microbial cells, resuspended in bead solution are added to a bead beating tube containing beads, followed by lysis solution. The principal is to lyse the microorganisms by a combination of heat, detergent, and mechanical force against specialized beads. The cellular components are lysed by mechanical action using a specially designed MO BIO Vortex Adapter on a standard vortex. From the lysed cells, the released DNA is bound to a silica Spin Filter. The filter is washed, and the DNA is recovered in certified DNA-free Tris buffer.

High Throughput Options

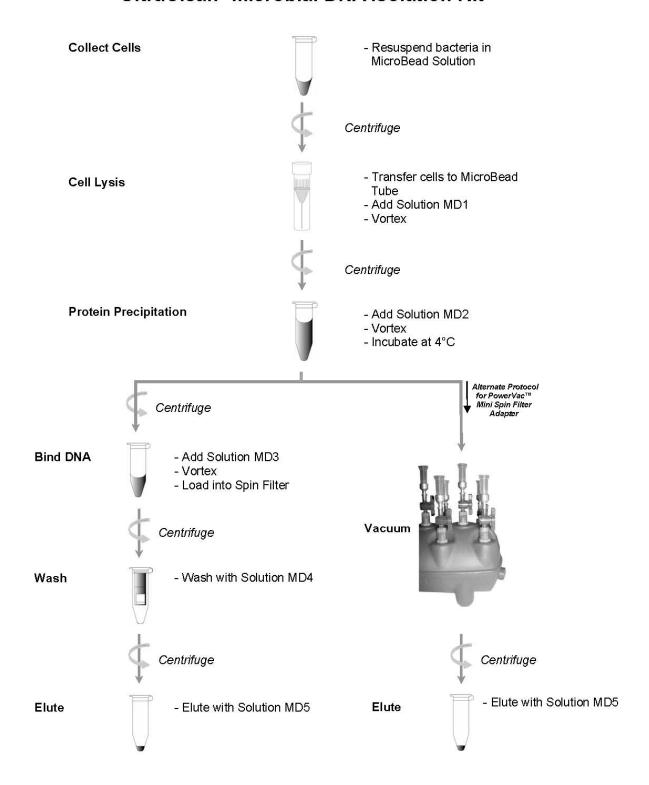
MO BIO offers a vacuum based protocol for faster processing without centrifugation for the DNA binding and column washing steps for Spin Filters. The MO BIO PowerVac[™] Manifold allows for processing of up to 20 spin filter preps at a time using the PowerVac[™] Mini Spin Filter Adapters. The UltraClean [®]-htp 96 Well Microbial DNA Isolation Kit is available for processing up to 2 x 96 samples using a centrifuge capable of spinning two 96 Well Blocks stacked (13 cm x 8 cm x 5.5 cm) at 2500 x g. For 96 well homogenization of bacteria, MO BIO offers the 96 Well Plate Shaker and Plate Adapter Set (MO BIO Catalog# 11996 & 11999, respectively.)

This kit is for research purposes only. Not for diagnostic use.

| Other Related Products | Catalog No. | Quantity |
|---|-------------|----------------------|
| UltraClean® Microbial RNA Isolation Kit | 15800-50 | 50 preps |
| | 15800-250 | 250 preps |
| UltraClean® PCR Clean-Up Kit | 12500-50 | 50 preps |
| · | 12500-100 | 100 preps |
| | 12500-250 | 250 preps |
| UltraClean®-htp 96 Well Microbial DNA | 10196-4 | 4 x 96 preps |
| Isolation Kit | 10196-12 | 12 x 96 preps |
| PowerVac™ Manifold | 11991 | 1 manifold |
| PowerVac™ Mini System | 11992 | 1 unit + 20 adapters |
| PowerVac™ Mini Spin Filter Adapters | 11992-10 | 10 adapters |
| · | 11992-20 | 20 adapters |



UltraClean® Microbial DNA Isolation Kit





Equipment Required

Microcentrifuge (10,000 x g) Pipettor (50 μ l – 200 μ l, 100 μ l – 1000 μ l) Vortex-Genie [®] 2 Vortex (MO BIO Catalog# 13111-V or 13111-V-220) Vortex Adapter (MO BIO Catalog# 13000-V1)

Reagents Required but not Included

100% ethanol (for the PowerVac™ Manifold protocol only)

Kit Contents

| | Kit Catalog # 12224-50 | | Kit Catalog # 122 | 24-250 |
|----------------------------------|------------------------|---------|-------------------|-----------|
| Component | Catalog # | Amount | Catalog # | Amount |
| MicroBead Tubes | 12224-50-BT | 50 | 12224-250-BT | 250 |
| (contain 250 mg MicroBeads) | | | | |
| MicroBead Solution | 12224-50-BS | 16.5 ml | 12224-250-BS | 80 ml |
| Solution MD1 | 12224-50-1 | 2.75 ml | 12224-250-1 | 15 ml |
| Solution MD2 | 12224-50-2 | 5.5 ml | 12224-250-2 | 30 ml |
| Solution MD3 | 12224-50-3 | 50 ml | 12224-250-3 | 250 ml |
| Solution MD4 | 12224-50-4 | 16.5 ml | 12224-250-4 | 3 x 32 ml |
| Solution MD5 | 12224-50-5 | 3 ml | 12224-250-5 | 15 ml |
| Spin Filters Units in 2 ml Tubes | 12224-50-SF | 50 | 12224-250-SF | 250 |
| 2 ml Collection Tubes | 12224-50-T | 200 | 12224-250-T | 1000 |

Kit Storage

Kit reagents and components should be stored at room temperature (15-30°C).

Precautions

Please wear gloves when using this product. Avoid all skin contact with kit reagents. In case of contact, wash thoroughly with water. Do not ingest. See Material Safety Data Sheets for emergency procedures in case of accidental ingestion or contact. All MSDS information is available upon request (760-929-9911) or at www.mobio.com. Reagents labeled flammable should be kept away from open flames and sparks.

WARNING: Solution MD4 contains ethanol, it is flammable. Do not use bleach to clean the inside of the PowerVac™ Manifold or to rinse the PowerVac™ Mini Spin Filter Adapters when attached to the manifold.

IMPORTANT NOTE FOR USE: Make sure the 2 ml MicroBead Tubes rotate freely in the centrifuge without rubbing. Do not spin the MicroBead Tubes in excess of 10,000 x g. Shake to mix Solution MD3 before use.



Experienced User Protocol

(If this is your first time using this kit please read the Detailed Protocol on the following page) Please wear gloves at all times

- 1. Add 1.8 ml of microbial (bacteria, yeast) culture to a **2 ml Collection Tube** (provided) and centrifuge at 10,000 x *g* for 30 seconds at room temperature. Decant the supernatant and spin the tubes at 10,000 x *g* for 30 seconds at room temperature and completely remove the media supernatant with a pipette tip. **NOTE**: Based on the type of microbial culture, it may be necessary to centrifuge longer than 30 seconds.
- 2. Resuspend the cell pellet in 300 μ l of **MicroBead Solution** and gently vortex to mix. Transfer resuspended cells to **MicroBead Tube**.
- 3. Check **Solution MD1**. If **Solution MD1** is precipitated, heat the solution at 60°C until the precipitate has dissolved. Add 50 µl of **Solution MD1** to the **MicroBead Tube**.
- 4. **Optional:** To increase yields, to minimize DNA shearing, or for difficult cells, see Alternative lysis methods in the "Hints & Troubleshooting Guide" section before continuing.
- 5. Secure **MicroBead Tubes** horizontally using the MO BIO Vortex Adapter tube holder for the vortex (MO BIO Catalog# 13000-V1) or secure tubes horizontally on a flat-bed vortex pad with tape. Vortex at maximum speed for 10 minutes. (See "Hints & Troubleshooting Guide" for less DNA shearing).
- 6. Make sure the 2 ml **MicroBead Tubes** rotate freely in the centrifuge without rubbing. Centrifuge the tubes at 10,000 x *g* for 30 seconds at room temperature. **CAUTION:** Be sure not to exceed 10,000 x *g* or tubes may break.
- 7. Transfer the supernatant to a clean **2 ml Collection Tube** (provided).
- 8. **NOTE**: Expect 300 to 350 μl of supernatant.
- 9. Add 100 μ l of **Solution MD2**, to the supernatant. Vortex for 5 seconds. Then incubate at 4°C for 5 minutes.
- 10. Centrifuge the tubes at room temperature for 1 minute at 10,000 x g.
- 11. Avoiding the pellet, transfer the entire volume of supernatant to a clean **2 ml Collection Tube** (provided). Expect approximately 450 µl in volume.
- 12. Shake to mix Solution MD3 before use. Add 900 μ l of **Solution MD3** to the supernatant and vortex for 5 seconds.
- 13. Load about 700 μ l into the **Spin Filter** and centrifuge at 10,000 x g for 30 seconds at room temperature. Discard the flow through, add the remaining supernatant to the **Spin Filter**, and centrifuge at 10,000 x g for 30 seconds at room temperature. **NOTE**: A total of 2 to 3 loads for each sample processed are required. Discard all flow through liquid.
- 14. Add 300 ul of **Solution MD4** and centrifuge at room temperature for 30 seconds at 10.000 x a.
- 15. Discard the flow through.
- 16. Centrifuge at room temperature for 1 minute at 10,000 x q.
- 17. Being careful not to splash liquid on the spin filter basket, place **Spin Filter** in a new **2 ml Collection Tube** (provided).
- 18. Add 50 µl of **Solution MD5** to the center of the white filter membrane.
- 19. Centrifuge at room temperature for 30 seconds at 10,000 x g.
- 20. Discard **Spin Filter**. The DNA in the tube is now ready for any downstream application. No further steps are required.

We recommend storing DNA frozen (-20°C). **Solution MD5** contains no EDTA.

Thank you for choosing the UltraClean® Microbial DNA Isolation Kit.



Detailed Protocol (Describes what is happening at each step) Please wear gloves at all times

1. Add 1.8 ml of microbial (bacteria, yeast) culture to a **2 ml Collection Tube** (provided) and centrifuge at 10,000 x *g* for 30 seconds at room temperature. Decant the supernatant and spin the tubes at 10,000 x *g* for 30 seconds at room temperature and completely remove the media supernatant with a pipette tip.

What's happening: This step concentrates and pellets the microbial cells. In some cases it may take longer to completely pellet the cells. It is important to pellet the cells completely and remove all the culture media in this step.

2. Resuspend the cell pellet in 300 μ l of **MicroBead Solution** and gently vortex to mix. Transfer resuspended cells to **MicroBead Tube**.

What's happening: The MicroBead Solution contains salts and a buffer which stabilizes and homogeneously disperses the microbial cells prior to lysis.

3. Check **Solution MD1**. If **Solution MD1** is precipitated, heat the solution at 60°C until the precipitate has dissolved. Add 50 µl of **Solution MD1** to the **MicroBead Tube**.

What's happening: Solution MD1 contains SDS and other disruption agents required for cell lysis. In addition to aiding in cell lysis, SDS is an anionic detergent that breaks down fatty acids and lipids associated with the cell membrane of several organisms. If it gets cold, it will precipitate. Heating at 60°C will dissolve the SDS and will not harm the SDS or the other disruption agents. In addition, Solution MD1 can be used while it is still warm.

4. **Optional:** To increase yields, to minimize DNA shearing, or for difficult cells, see Alternative lysis methods in the "Hints & Troubleshooting Guide" section before continuing.

What's happening: This optional step can lead to better performance in some cases. We recommend using only one of these methods for any individual prep.

5. Secure **MicroBead Tubes** horizontally using the MO BIO Vortex Adapter tube holder for the vortex (MO BIO Catalog# 13000-V1) or secure tubes horizontally on a flat-bed vortex pad with tape. Vortex at maximum speed for 10 minutes. (See "Hints & Troubleshooting Guide" for less DNA shearing).

What's happening: This step creates the combined chemical/ mechanical lysis conditions required to release desired nucleic acids from microbial cells. Many cell types will not lyse without this chemically enhanced bead beating process. The vortex action is typically all that is required, however, more robust bead beaters may also be used. In most cases the times may be shorter with other devices but you may run the risk of increased DNA shearing. This process is compatible with fast prep machines.

6. Make sure the 2 ml **MicroBead Tubes** rotate freely in the centrifuge without rubbing. Centrifuge the tubes at 10,000 x g for 30 seconds at room temperature. **CAUTION:** Be sure not to exceed 10,000 x g or tubes may break.

What's happening: The cell debris is sent to the bottom of the tube while DNA is remains in the supernatant.

7. Transfer the supernatant to a clean 2 ml Collection Tube (provided).



8. **NOTE**: Expect 300 to 350 μl of supernatant.

What's happening: The volume to expect will vary depending on the size of the original cell pellet from step 1.

- Add 100 μl of Solution MD2, to the supernatant. Vortex for 5 seconds. Then incubate at 4°C for 5 minutes.
- 10. Centrifuge the tubes at room temperature for 1 minute at 10,000 x g.

What's happening: Solution MD2 contains a reagent to precipitate non-DNA organic and inorganic material including cell debris and proteins. It is important to remove contaminating organic and inorganic matter that may reduce DNA purity and inhibit downstream DNA applications.

11. Avoiding the pellet, transfer the entire volume of supernatant to a clean **2 ml Collection Tube** (provided). Expect approximately 450 μl in volume.

What's happening: The pellet at this point contains non-DNA organic and inorganic materials, including cell debris and proteins. For the best DNA quality and yield, avoid transferring any of the pellet.

12. Shake to mix Solution MD3 before use. Add 900 μ l of **Solution MD3** to the supernatant and vortex for 5 seconds.

What's happening: Solution MD3 is a highly concentrated salt solution. It sets up the high salt condition necessary to bind DNA to the Spin Filter membrane in the following step.

13. Load about 700 μl into the **Spin Filter** and centrifuge at 10,000 x *g* for 30 seconds at room temperature. Discard the flow through, add the remaining supernatant to the **Spin Filter**, and centrifuge at 10,000 x *g* for 30 seconds at room temperature. **NOTE**: A total of 2 to 3 loads for each sample processed are required. Discard all flow through liquid.

What's happening: DNA is selectively bound to the silica membrane in the Spin Filter device. Contaminants pass through the filter membrane, leaving only the DNA bound to the membrane.

14. Add 300 ul of **Solution MD4** and centrifuge at room temperature for 30 seconds at 10,000 x q.

What's happening: Solution MD4 is an ethanol based wash solution used to further clean the DNA that is bound to the silica filter membrane in the Spin Filter. This wash solution removes residues of salt, and other contaminants while allowing the DNA to stay bound to the silica membrane.

15. Discard the flow through.

What's happening: This flow through is waste containing ethanol wash solution and contaminants that did not bind to the silica Spin Filter membrane.

16. Centrifuge at room temperature for 1 minute at 10,000 x g.

What's happening: This step removes residual Solution MD4 (ethanol wash solution). It is critical to remove all traces of wash solution because it can interfere with down stream DNA applications.

17. Being careful not to splash liquid on the spin filter basket, place **Spin Filter** in a new **2 ml Collection Tube** (provided).



What's happening: It is important to avoid any traces of the ethanol based wash solution.

18. Add 50 μ l of **Solution MD5** to the center of the white filter membrane.

What's happening: Placing the Solution MD5 (elution buffer) in the center of the small white membrane will make sure the entire membrane is wetted. This will result in more efficient release of bound DNA

19. Centrifuge at room temperature for 30 seconds at 10,000 x g.

What's happening: As the Solution MD5 (elution buffer) passes through the silica membrane, DNA is released, and it flows through the membrane, and into the Collection Tube. The DNA is released because it can only bind to the silica Spin Filter membrane in the presence of salt. Solution MD5 is 10mM Tris pH 8 and does not contain salt.

20. Discard **Spin Filter**. DNA in the tube is now ready for any downstream application. No further steps are required.

We recommend storing DNA frozen (-20°C). **Solution MD5** contains no EDTA.

Thank you for choosing the UltraClean® Microbial DNA Isolation Kit.



Vacuum Protocol using the PowerVac™ Manifold Please wear gloves at all times

For each sample lysate, use one Spin Filter column. Keep the Spin Filter in the attached 2 ml Collection Tube and continue using the Collection Tube as a Spin Filter holder until needed for the Vacuum Manifold Protocol. Label each Collection Tube top and Spin Filter column to maintain sample identity. If the Spin Filter becomes clogged during the vacuum procedure, you can switch to the procedure for purification of the DNA by centrifugation.

You will need to provide 100% ethanol for step 4 of this protocol

For each prep, attach one aluminum PowerVac[™] Mini Spin Filter Adapter (MO BIO Catalog# 11992-10 or 11992-20) into the Luer-Lok® fitting of one port in the manifold. Gently press a Spin Filter column into the PowerVac[™] Mini Spin Filter Adapter until snugly in place. Ensure that all unused ports of the vacuum manifold are closed.

Note: Aluminum PowerVac™ Mini Spin Filter Adapters are reusable.

- 2. Transfer 650 µl of prepared sample lysate (from step 12) to the **Spin Filter column**.
- 3. Turn on the vacuum source and open the stopcock of the port. Hold the tube in place when opening the stopcock to keep the spin filter steady. Allow the lysate to pass through the **Spin Filter column**. After the lysate has passed through the column completely, load again with the next 650 µl of lysate. Continue until all of the lysate has been loaded onto the **Spin Filter column**. Close the one-way Luer-Lok® stopcock of that port.

Note: If Spin Filter Columns are filtering slowly, close the ports to samples that have completed filtering to increase the pressure to the other columns.

- 4. Load 800 μ l of 100% ethanol into the Spin Filter so that it completely fills the column. Open the stopcock while holding the column steady. Allow the ethanol to pass through the column completely. Close the stopcock.
- 5. Add 300 µl of **Solution MD4** to each Spin Filter. Open the Luer-Lok® stopcock and apply a vacuum until **Solution MD4** has passed through the Spin Filter completely. Continue to pull a vacuum for another minute to dry the membrane. Close each port.
- 6. Turn off the vacuum source and open an unused port to vent the manifold. If all 20 ports are in use, break the vacuum at the source. Make certain that all vacuum pressure is released before performing the next step. It is important to turn off the vacuum at the source to prevent backflow into the columns.
- 7. Remove the **Spin Filter column** and place in the original labeled **2 ml Collection Tube**. Place into the centrifuge and spin at $13,000 \times g$ for 1 minute to completely dry the membrane.
- 8. Transfer the **Spin Filter column** to a new **2 ml Collection Tube** and add 50 μ l of **Solution MD5** to the center of the white filter membrane. Alternatively, sterile DNA-Free PCR Grade Water may be used for elution from the silica **Spin Filter** membrane at this step (MO BIO Catalog# 17000-10).
- 9. Centrifuge at room temperature for 30 seconds at 10,000 x g.



10. Discard the **Spin Filter column**. The DNA in the tube is now ready for any downstream application. No further steps are required.

We recommend storing DNA frozen (-20° to -80°C). **Solution MD5** contains no EDTA. To concentrate the DNA see the Hints & Troubleshooting Guide.

Thank you for choosing the UltraClean® Microbial DNA Isolation Kit.



Hints and Troubleshooting Guide

Alternative Lysis Methods (We recommend using only one of these methods for any individual prep.)

- **To increase yields**: Heating can aid in lysis for some organisms and it can lead to increased yields. Heat preps at 65°C for 10 minutes and continue with step 5.
- For less DNA shearing: We recommend heating the preps at 65°C for 10 minutes with occasional bump vortexing for a few seconds every 2-3 minutes. Skip step 5 and go to step 6. This helps prevent unwanted damage to large DNA. This procedure will reduce DNA shearing and at the same time can increase the yield of total DNA for some organisms.
- If cells are difficult to lyse: Heat the preps at 70°C for 10 minutes. Follow by continuing with the protocol at step 5.

Concentrating the DNA

The final volume of eluted DNA will be 50 μ l. The DNA may be concentrated by adding 5 μ l of 5M NaCl and inverting 3-5 times to mix. Next, add 100 μ l of 100% cold ethanol and invert 3-5 times to mix. Incubate at -20°C for 30 minutes and centrifuge at 10,000 x g for 15 minutes at room temperature. Decant all liquid. Remove residual ethanol in a speed vac or dessicator or air dry. Resuspend precipitated DNA in sterile water or Solution MD5 (10 mM Tris).

DNA Floats Out of Well When Loaded on a Gel

This usually occurs because residual Solution MD4 remains in the final sample. Prevent this by being careful in step 17 not to transfer liquid onto the bottom of the spin filter basket. Ethanol precipitation (described in "Concentrating the DNA") is the best way to remove residual Solution MD4.

Storing DNA

DNA is eluted in Solution MD5 (10 mM Tris) and must be stored at -20°C to -80°C to prevent degradation. For long term storage, we recommend aliquoting DNA into appropriate volumes and store at -80°C. DNA can be eluted in TE without loss, but the EDTA may inhibit downstream reactions such as PCR and automated sequencing. DNA may also be eluted with sterile DNA-Free PCR Grade Water (MO BIO Catalog# 17000-10).

Cleaning of the PowerVac™ Mini Spin Filter Adapters

It is recommended to rinse the PowerVac[™] Mini Spin Filter Adapters promptly after use to avoid salt build up. To clean the PowerVac[™] Mini Spin Filter Adapters, rinse each adapter with DI water followed by 70% ethanol and flush into the manifold base. Alternatively, remove the adapters and wash in laboratory detergent and DI water. PowerVac[™] Mini Spin Filter Adapters may be autoclaved.

Do not use bleach to clean the PowerVac[™] Mini Spin Filter Adapters while attached to the PowerVac[™] Manifold. Bleach should never be mixed with solutions containing guanidine and should not be used to clean the PowerVac[™] Manifold. For more information on cleaning the PowerVac[™] Manifold, please refer to the PowerVac[™] Manifold manual.



Contact Information

Technical Support:

Phone MO BIO Laboratories, Inc. Toll Free 800-606-6246, or 760-929-9911

Email: technical@mobio.com

Fax: 760-929-0109

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For the distributor nearest you, visit our web site at www.mobio.com/distributors



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|--|-------------|------------------------|
| DNA Purification and Gel Extraction | Catalog No. | Quantity |
| PowerClean® DNA Clean-Up Kit | 12877-50 | 50 preps |
| UltraClean® 15 DNA Purification Kit | 12100-300 | 300 preps |
| UltraClean® PCR Clean-Up Kit | 12500-50 | 50 preps |
| | 12500-100 | 100 preps |
| | 12500-250 | 250 preps |
| UltraClean®-htp 96 Well PCR Clean- | 12596-4 | 4 x 96 preps |
| Up Kit | 12596-12 | 12 x 96 preps |
| UltraClean® GelSpin® DNA | 12400-50 | 50 preps |
| Extraction Kit | 12400-100 | 100 preps |
| | 12400-250 | 250 preps |
| Plasmid DNA Isolation | Catalog No. | Quantity |
| UltraClean® 6 Minute Mini Plasmid | 12300-50 | 50 preps |
| Prep Kit | 12300-100 | 100 preps |
| • | 12300-250 | 250 preps |
| UltraClean® Standard Mini Plasmid | 12301-50 | 50 preps |
| Prep Kit | 12301-100 | 100 preps |
| | 12301-250 | 250 preps |
| UltraClean®-htp 96 Well Plasmid Prep | 12396-4 | 4 x 96 preps |
| Kit | 12396-12 | 12 x 96 preps |
| UltraClean® Midi Plasmid Prep Kit | 12700-20 | 20 preps |
| _ | 12700-50 | 50 preps |
| UltraClean® Maxi Plasmid Prep Kit | 12600-10 | 10 preps |
| | 12600-20 | 20 preps |
| UltraClean® Endotoxin-Free Mini | 12311-100 | 100 preps |
| Plasmid Prep Kit | 12311-250 | 250 preps |
| UltraClean® Endotoxin-Free Midi Plasmid Prep Kit | 12711-10 | 10 preps |
| UltraClean® Endotoxin-Free Maxi Plasmid Prep Kit | 12611-10 | 10 preps |
| UltraClean® Endotoxin Removal Kit | 12615 | 1 kit |
| UltraClean® Endotoxin-Free Ethanol Precipitation Kit | 12616 | 1 kit |
| UltraClean® Endotoxin Removal Reagent | 12625-25 | 25 ml |
| Endotoxin-Free Sodium Chloride | 12626-15 | 15 ml |
| Endotoxin-Free Centrifuge Tubes | 12617-100 | 100 each/2 ml tubes |
| | 12618-50 | 50 each/15 ml tubes |
| | 12619-25 | 25 each/50 ml tubes |
| | | |
| RNA Isolation | Catalog No. | Quantity |
| PowerBiofilm™ RNA Isolation Kit | 25000-50 | 50 preps |
| LifeGuard™ Soil Stabilization Solution | 12868-10 | 10 ml |
| Listada Goil Glabilization Goiution | 12868-100 | 100 ml |
| | 12868-1000 | 1 L |
| | 12868-7500 | 7.5 L |
| On-Spin Column DNase I Kit (RNase- | 15100-50 | 50 preps |
| Free) | | |
| Bi Ostic® Stabilized Blood RNA | 12231-20 | 20 preps |
| Isolation Kit | 12231-50 | 50 preps |
| Dio (CD III III III III | 12231-100 | 100 preps |
| Bi Ostic® Blood Total RNA Isolation | 12230-20 | 20 preps |
| Kit RNA Dougresil® DNA Flution | 12230-50 | 50 preps |
| RNA PowerSoil® DNA Elution Accessory Kit | 12867-25 | 25 preps |
| RNA PowerSoil® Total RNA Isolation Kit | 12866-25 | 25 preps |
| UltraClean® Microbial RNA Isolation | 15800-50 | 50 preps |
| Kit | 15800-250 | 250 preps |

| RNA Isolation Continued | Catalog No. | Quantity |
|--|-----------------------|-----------------------|
| UltraClean® Tissue & Cells RNA | 15000-50 | 50 preps |
| Isolation Kit | 15000-250 | 250 preps |
| UltraClean® Plant RNA Isolation Kit | 13300-20 | 20 preps |
| | 13300-50 | 50 preps |
| | | |
| Genomic DNA Isolation | Catalog No. | Quantity |
| PowerBiofilm™ DNA Isolation Kit | 24000-50 | 50 preps |
| PowerFood ™ Microbial DNA Isolation | 21000-50 | 50 preps |
| Kit | 21000-100 | 100 preps |
| Bi Ostic® Bacteremia DNA Isolation Kit | 12240-50 | 50 preps |
| Bi Ostic® FFPE Tissue DNA Isolation | 12250-50 | 50 preps |
| Kit | | |
| Bi Ostic® Paraffin Removal Reagent | 12251-50 | 2 x 25 ml |
| | | |
| PowerMax® Soil DNA Isolation Kit | 12988-10 | 10 preps |
| PowerSoil® DNA Isolation Kit | 12888-50 | 50 preps |
| | 12888-100 | 100 preps |
| PowerSoil®-htp 96 Well Soil DNA | 12955-4 | 4 x 96 preps |
| Isolation Kit | 12955-12 | 12 x 96 prep |
| UltraClean® Soil DNA Isolation Kit | 12800-50 | 50 preps |
| | 12800-100 | 100 preps |
| UltraClean®-htp 96 Well Soil DNA | 12896-4 | 4 x 96 preps |
| Isolation Kit | 12896-12 | 12 x 96 prep |
| UltraClean® Mega Soil DNA Isolation Kit | 12900-10 | 10 preps |
| PowerClean® DNA Clean-Up Kit | 12877-50 | 50 preps |
| UltraClean® Fecal DNA Isolation Kit | 12811-50 | 50 preps |
| Chiadioane i doai Dian Isolation Mil | 12811-100 | 100 preps |
| PowerMicrobial® Midi DNA Isolation Kit | 12225-25 | 25 preps |
| PowerMicrobial® Maxi DNA Isolation Kit | 12226-25 | 25 preps |
| UltraClean® Microbial DNA Isolation | 12224-50 | 50 preps |
| Kit | 12224-250 | 250 preps |
| UltraClean®-htp 96 Well Microbial | 10196-4 | 4 x 96 preps |
| DNA Isolation Kit | 10196-12 | 12 x 96 prep |
| PowerPlant® DNA Isolation Kit | 13200-50 | 50 preps |
| | 13200-100 | 100 preps |
| UltraClean® Plant DNA Isolation Kit | 13000-50 13000-250 | 50 preps 250 preps |
| UltraClean®-htp 96 Well Plant DNA Isolation Kit | 13096-4 13096-12 | 4 x 96 preps |
| UltraClean® Tissue & Cells DNA | 12334-50 | 50 preps |
| Isolation Kit | 12334-250 | 250 preps |
| UltraClean®-htp 96 Well Tissue DNA | 12996-4 | 4 x 96 preps |
| Isolation Kit | 12996-12 | 12 x 96 prep |
| UltraClean® Blood DNA Isolation Kit (Non-Spin) | 12000-100 | 100 preps |
| UltraClean® Blood DNA Isolation Kit (Processes 1,000 ml of Blood) | 12000-1000 | 1 kit |
| UltraClean® Blood DNA Isolation Kit Plus RNase (Processes 1,000 ml of Blood) | 12002-1000 | 1 kit |



| Genomic DNA Isolation | | |
|---|--|---|
| Continued | Catalog No. | Quantity |
| UltraClean® BloodSpin® DNA Isolation Kit | 12200-50 12200-250 | 50 preps 250 preps |
| UltraClean®-htp 96 Well BloodSpin® | 12296-4 | 4 x 96 preps |
| DNA Isolation Kit | 12296-12 | 12 x 96 preps |
| UltraClean® Forensic DNA Isolation | 14000-10 | 10 isolations |
| Kit | 14000-20 | 20 isolations |
| PowerWater® DNA Isolation Kit | | 50 preps |
| | 14900-50-NF | (No filters) |
| | 14900-50-22 | (0.22 µm) |
| | 14900-50-45 | (0.45 µm) |
| | 4.4000 400 NE | 100 preps |
| | 14900-100-NF | (No filters) |
| | 14900-100-22 14900-100-45 | (0.22 μm) (0.45 μm) |
| RapidWater™ DNA Isolation Kit | 14300-100-43 | 50 preps |
| rapidivator Brivi Isolation (tit | 14810-50-NF | (No filters) |
| | 14810-50-22 | (0.22 µm) |
| | 14810-50-45 | (0.45 µm) |
| | | 100 preps |
| | 14810-100-NF | (No filters) |
| | 14810-100-22 | (0.22 µm) |
| | 14810-100-45 | (0.45 µm) |
| UltraClean® Water DNA Isolation Kit | 14800-10 | 10 preps |
| (0.45µm filters) | 14800-25 | 25 preps |
| UltraClean® Water DNA Isolation Kit | 14880-10 | 10 preps |
| (0.22 µm filters) | 14880-25 | 25 preps |
| (0.22 µm mters) | 11000 20 | 20 01000 |
| UltraClean® Water DNA Isolation Kit | 14800-10-NF | 10 preps |
| (No filters) | 14800-25-NF | 25 preps |
| | | |
| Microbiological Cultura Madia | Catalan Na | Overetite. |
| Microbiological Culture Media TB DRY® Powder Growth Media | Catalog No. 12105-05 | Quantity 500 q |
| 16 DK 1 @ Fowder Glowii i wiedia | 12105-05 | 1 kg |
| | 12105-5 | 5 kg |
| LB Broth Powder Growth Media, pH | 12106-05 | 500 g |
| 7 | 12106-1 | 1 kg |
| | 12106-5 | 5 kg |
| LB Agar Powder Growth Media, pH 7 | 12107-05 | 500 g |
| | 12107-1 | 1 kg |
| | 12107-5 | 5 kg |
| LB Broth (Lennox) Powder Growth | 12108-05 | 500 g |
| Media, pH 7 | 12108-1 12108-5 | 1 kg 5 kg |
| LB Agar (Lennox) Powder Growth | 12108-5 | 5 kg 500 g |
| Media, pH 7 | 12109-03 | 1 kg |
| modia, pri i | 12109-1 | 5 kg |
| Soybean-Casein Digest Medium | 12114-05 | 500 g |
| (TSB), USP | 12114-1 | 1 kg |
| | 12114-5 | 5 kg |
| | I | |
| | | |
| Soybean-Casein Digest Agar | 12115-05 | 500 g |
| Soybean-Casein Digest Agar Medium (TSA), USP | 12115-1 | 1 kg |
| Soybean-Casein Digest Agar Medium (TSA), USP | | |
| Soybean-Casein Digest Agar Medium (TSA), USP | 12115-1 | 1 kg |
| Soybean-Casein Digest Agar Medium (TSA), USP | 12115-1 | 1 kg |
| Medium (TSA), USP | 12115-1 12115-5 12110-05 12110-1 | 1 kg 5 kg 500 g 1 kg |
| Medium (TSA), USP Yeast Extract | 12115-1 12115-5 12110-05 | 1 kg 5 kg 500 g 1 kg 5 kg |
| Medium (TSA), USP | 12115-1 12115-5 12110-05 12110-1 12110-5 12111-05 | 1 kg 5 kg 500 g 1 kg 5 kg 500 g |
| Medium (TSA), USP Yeast Extract | 12115-1 12115-5 12110-05 12110-1 12110-5 12111-05 12111-1 | 1 kg 5 kg 500 g 1 kg 5 kg 500 g 1 kg |
| Medium (TSA), USP Yeast Extract Tryptone | 12115-1 12115-5 12110-05 12110-1 12110-5 12111-05 12111-1 12111-5 | 1 kg 5 kg 500 g 1 kg 5 kg 500 g 1 kg 5 kg |
| Medium (TSA), USP Yeast Extract | 12115-1 12115-5 12110-05 12110-1 12110-5 12111-05 12111-1 12111-5 12112-05 | 1 kg 5 kg 500 g 1 kg 5 kg 500 g 1 kg 5 kg 500 g |
| Medium (TSA), USP Yeast Extract Tryptone | 12115-1 12115-5 12110-05 12110-1 12110-5 12111-05 12111-1 12111-5 | 1 kg 5 kg 500 g 1 kg 5 kg 500 g 1 kg 5 kg |

| Other Reagents and Lab | | |
|--|-------------------------|--------------------------------|
| Accessories | Catalog No. | Quantity |
| 20 bp DNA Ladder | 17020-40 | 40 μg |
| | | |
| 100 bp DNA Ladder | 17100-40 | 40 μg |
| 1 kb DNA Ladder | 17200-100 | 100 µg |
| UltraClean® Agarose, Molecular | 15003-50 | 50 g |
| Biology Grade | 15003-100 15003-500 | 100 g 500 g |
| | 15003-300 | 1 kg |
| | | 19 |
| | | |
| Lillara Classica MC O A careas | 15515-50 | 50 g |
| UltraClean® MS-8 Agarose | 15515-100 | 100 g |
| | 15515-500 | 500 g |
| | | |
| | | |
| | | |
| UltraClean® Forensic Agarose | 15505-50 | 50 g |
| Oliracicane i orensio rigarose | 15505-100 | 100 g |
| | 15505-500 | 500 g |
| UltraClean® Low Melt Agarose | 15005-50 | 50 g |
| | 15005-100 15005-500 | 100 g 500 g |
| UltraClean® Low Melt Sieve Agarose | 15003-500 | 50 g |
| , and the second | 15004-100 | 100 g |
| Establisher Borowide Calastia | 15004-500 | 500 g |
| Ethidium Bromide Solution | 15006-1 15006-10 | 1 ml 10 ml |
| Ethidium Bromide Destaining Tea | 15007-25 | 25 bags |
| Bags | | ŭ |
| Bromophenol Blue Gel Loading | 15008-1 | 1 ml |
| Buffer | 15008-5 | 5 x 1 ml |
| Bromophenol Blue/Xylene Cyanol | 15009-1 | 1 ml |
| Gel Loading Buffer | 15009-5 | 5 x 1 ml |
| TAE DUKET FOY (Trip posteto FDTA) | 45004.400 | 400 |
| TAE Buffer, 50X (Tris-acetate-EDTA) | 15001-100 15001-500 | 100 ml 500 ml |
| | 15001-1000 | 1 liter |
| TBE Buffer, 10X (Tris-borate-EDTA) | 15002-100 | 100 ml |
| | 15002-500 15002-1000 | 500 ml 1 liter |
| RNase-Free Gloves | 1555-XS | bag of 100 |
| | 1555-S | bag of 100 |
| | 1555-M | bag of 100 |
| UltraClean® Lab Cleaner | 1555-L 12095-250 | bag of 100 250 ml |
| S. Colouro Las Sidario | .2000 200 | squeeze bottle |
| | 12095-500 | 500 ml spray |
| | 12095-1000 | bottle 1 liter bottle |
| KAPA PROBE FAST qPCR Kits | 51220-100 | 100 reactions |
| | 51220-500 | 500 reactions |
| WARA 6V/PRO 5455 | 51220-1000 | 1000 reactions |
| KAPA SYBR® FAST Universal 2X gPCR Master Mix | 51230-100 51230-500 | 100 reactions 500 reactions |
| q. Or master mix | 51230-300 | 1000 reactions |
| KAPA2G Robust HotStart ReadyMix | 51240-100 | 100 reactions |
| | 51240-500 | 500 reactions |
| l . | 1 | 1 |



| Other Reagents and Lab | | |
|---|------------------------|--------------------------------|
| AccessoriesContinued | Catalog No. | Quantity |
| KAPA HiFi HotStart ReadyMix | 51250-100 | 100 reactions |
| , | 51250-500 | 500 reactions |
| KAPA2G FAST HotStart DNA | 51260-100 | 100 reactions |
| Polymerase with dNTPs | 51260-250 | 250 reactions |
| | 51260-500 | 500 reactions |
| KAPA2G FAST HotStart ReadyMix | 51270-100 | 100 reactions |
| KARAL B. H. O. (DNA | 51270-500 | 500 reactions |
| KAPA Long Range HotStart DNA Polymerase with dNTPs | 51280-100 51280-250 | 100 reactions 250 reactions |
| Folymerase with diviris | 51280-500 | 500 reactions |
| KAPA Taq Polymerase ReadyMix | 51290-250 | 250 reactions |
| O IT THE DAMA D. I | 1001.050 | 050 " |
| OmniTaq™ DNA Polymerase | 1224-250 | 250 reactions |
| Enzyme OmniTaq™ DNA Polymerase 2x | 1226-250 | (10 U/µI) 250 reactions |
| OmniTaq™ DNA Polymerase 2x Master Mix | 1220-230 | (5 x 1.25 |
| IVIASIEI IVIIX | | ml/tube) |
| Omni KlenTaq™ DNA Polymerase | 1225-250 | 250 reactions |
| Enzyme | 1220 200 | (25 U/µI) |
| Omni KlenTaq™ DNA Polymerase 2x | 1227-250 | 250 reactions |
| Master Mix | | (5 x 1.25 |
| | | ml/tube) |
| DNase (RNase-Free) | 15600-5 | 5 mg |
| Divase (Midse-1186) | 15601-100 | 2500 units |
| Proteinase K | 1223-100 | 100 mg |
| 1 Totolilase IX | 1222-2 | 2 ml (20 |
| | | mg/ml) |
| | | , |
| Ribonuclease A (25 mg/ml) | 1202-1 | 1 ml |
| | 1202-5 | 5 ml |
| PCR Water | 17000-1 | 1 ml |
| | 17000-5 | 5 x 1 ml |
| | 17000-10 17000-11 | 10 x 1 ml 10 ml bottle |
| Molecular Biology Grade Water | 17012-200 | 200 ml |
| Wildlectial Biology Grade Water | 17012-200 | 5 x 200 ml |
| DEPC Treated Water | 17011-200 | 200 ml |
| | 17011-5200 | 5 x 200 ml |
| Endotoxin-Free Water | 17013-10 | 10 ml |
| | 17013-50 | 50 ml |
| | 17013-100 | 100 ml |
| | 17013-500 | 500 ml |
| | | |
| Instrumentation and Accessories | Catalog No. | Quantity |
| PowerLyzer™ 24 Bench Top Bead- | 13155 | 1 unit |
| Based Homogenizer (110/220V) | 10100 | 1 UIII |
| PowerLyzer™ Tube Holder | 13156 | 1 unit |
| PowerLyzer™ Tube Holder Stand | 13157 | 1 unit |
| | | |
| PowerVac™ Mini System | 11992 | 1 unit + 20 adapters |
| PowerVac™ Manifold | 11991 | 1 unit |
| | | |
| PowerVac™ Mini Spin Filter | 11992-10 | 10 adapters |
| Adapters | 11992-20 | 20 adapters |
| | | |
| | | |
| Ceramic Bead Tubes, 1.4 mm | 13113-50 | 50 bead tubes |
| | 13114-50 | 50 bead tubes |
| Ceramic Bead Tubes, 2.8 mm | | |

| Instrumentation and | Cotolog No | Quantity |
|--|---------------------------|---------------------------|
| Accessories Continued Glass Bead Tubes, 0.5 mm | Catalog No. 13116-50 | Quantity 50 bead tubes |
| Class Boad Tables, c.s IIIII | 1011000 | oo boaa taboo |
| Glass Bead Tubes, 0.1 mm | 13118-50 | 50 bead tubes |
| | | |
| Metal Bead Tubes, 2.38 mm | 13117-50 | 50 bead tubes |
| Motal Boda Taboo, 2.50 mm | 10111 00 | oo boaa taboo |
| 2.0 ml Tough Tubes with Cap | 13119-500 | 500 |
| | 13119-1000 | 1000 |
| Carbide Bead Tubes, 0.25 mm | 13121-50 | 50 x 0.5 ml |
| - Carbiae Dead Fabes, 6:20 | 10.2.00 | tubes |
| Garnet Bead Tubes, 0.15 mm | 13122-50 | 50 x 0.5 ml |
| Operat Dead Takes 0.70 mag | 40400 50 | tubes |
| Garnet Bead Tubes, 0.70 mm | 13123-50 | 50 x 2 ml tubes |
| | | tubes |
| Garnet + 1/4 Ceramic 15 ml Bead | 13134-50 | 50 tubes |
| Tubes, 0.70 mm | | |
| Garnet + ¼ Ceramic 50 ml Bead | 13144-10 | 10 tubes |
| Tubes, 0.70 mm ` | 13144-50 13144-100 | 50 tubes 100 tubes |
| | 13144-500 | 500 tubes |
| Glass 15 ml Bead Tubes, 0.1 mm | 13135-50 | 50 tubes |
| Glado 10 III Boda 1 aboo, 0.1 IIIIII | 10100 00 | 00 14500 |
| Glass 50 ml Bead Tubes, 0.1 mm | 13145-10 | 10 tubes |
| | 13145-50 | 50 tubes |
| | 13145-100 | 100 tubes |
| | 13145-500 | 500 tubes |
| Glass 15 ml Bead Tubes, 1.0 mm | 13136-50 | 50 tubes |
| Ceramic 15 ml Bead Tubes, 1.4 mm | 13137-50 | 50 tubes |
| Column To III Bodd Tuboo, II IIIIII | 10101 00 | 00 10000 |
| | | |
| | | |
| Ceramic 50 ml Bead Tubes, 1.4 mm | 13147-10 | 10 tubes |
| M / LEO LD LT L OOG | 13147-50 | 50 tubes |
| Metal 50 ml Bead Tubes, 2.38 mm | 13149-10 13149-50 | 10 tubes 50 tubes |
| PowerMix 15 ml Bead Tubes | 13138-50 | 50 tubes |
| 1 OWERWIX 13 IIII Dead Tubes | 13130-30 | 30 tubes |
| | | |
| | | |
| PowerMix 50 ml Bead Tubes | 13148-10 | 10 tubes |
| | 13148-50 | 50 tubes |
| | | |
| 2 ml Collection Tubes | 1200-100-T | 100 tubes |
| | 1200-150-T | 150 tubes |
| | 1200-250-T | 250 tubes |
| 2 ml Screw Cap Tubes | 12800-200-E | 200 tubes & |
| 45 and Online tion Tables | 40700 T | caps |
| 15 ml Collection Tubes | 12700-T | 25 tubes |
| 50 ml Centrifuge Tubes | 12600-T | 25 tubes |
| Spin Filters (in 1.9 ml tubes) | 1200-50-SF | 50 filters |
| | 1200-30-31 1200-100-SF | 100 filters |
| | 1200-250-SF | 250 filters |
| Endotoxin-Free Centrifuge Tubes | 12617-100 | 100 each/2 ml |
| | | tubes |
| | 12618-50 | 50 each/15 ml |
| | 12610.25 | tubes |
| | 12619-25 | 25 each/50 ml tubes |
| 15 ml Midi Spin Filters | 12700-SF | 25 spin filters |
| 15 th that opin i more | 12,000 | _0 op intoio |
| | | |



| Vortex-Genie® 2 Vortex (120V) 1311 Vortex-Genie® 2 Vortex (220V) 1311 Vortex Adapter, holds 12 (1.5-2.0 ml) 1300 tubes | 1-V 220 1 unit 100-V1 1 unit |
|--|------------------------------|
| Vortex-Genie® 2 Vortex (220V) 1311 Vortex Adapter, holds 12 (1.5-2.0 ml) tubes | 1-V-220 1 unit |
| Vortex Adapter, holds 12 (1.5-2.0 ml) 1300 tubes | |
| tubes | 70-V1 1 d1iit |
| Vortex Adapter, holds 6 (5 ml) tubes 1300 | |
| , , , , , | 00-V1-5 1 unit |
| Vortex Adapter, holds 4 (15 ml) tubes 1300 | 00-V1-15 1 unit |
| Vortex Adapter, holds 2 (50 ml) tubes 1300 | 00-V1-50 1 unit |
| Vortex Adapter, holds 24 (1.5-2.0 ml) 1300 tubes | 00-V1 <i>-</i> 24 1 unit |
| BagMixer® 400 VW 2311 | 2 1 unit |
| BagFilter® 400 P 2311 | 3-500 Box of 500 |
| BagPage® 400 2311 | 4-500 Box of 500 |
| Whirl-Pak® Collection Bag, Small (532 ml) | 0-500 500 bags |
| Whirl-Pak® Collection Bag, Medium (1,627 ml) 2321 | 1-500 500 bags |
| Whirl-Pak® Collection Bag, Large (3,637 ml) | 2-250 250 bags |
| Whirl-Pak® Stand up Bag, Small 2322 (118 ml) | 20-500 500 bags |
| Whirl-Pak® Stand up Bag, Medium (532 ml) 2322 | 21-500 500 bags |
| Whirl-Pak® Stand up Bag, Large 2322 (1,242 ml) | 22-250 250 bags |
| Whirl-Pak® Stand up Bag, Extra- Large (2,041 ml) | 23-250 250 bags |
| Whirl-Pak® Scoop Bag, 60 ml 2324 | 10-50 50 bags |
| Anti-Static Funnels, Micro 2330 | 01-96 Pack of 96 |
| Anti-Static Funnels, Small 2330 | 02-50 Pack of 50 |
| Anti-Static Funnels, Medium 2330 | 03-50 Pack of 50 |
| Anti-Static Funnels, Large 2330 | 04-20 Pack of 20 |

| Instrumentation and Accessories Continued | Catalog No. | Quantity |
|---|----------------|---------------------------------|
| Mini Horizontal Gel System | 16001 | 1 each |
| Mini Horizontal Gel Caster, 3 place | 16003 | 1 each |
| Mini Horizontal Gel Tray | 16004 | 1 each |
| Deliverabe pate Cingle sided Comb | 10005 | 4 |
| Polycarbonate Single-sided Comb | 16005 16006 | 1 mm x 3 well 1 mm x 8 well |
| | 16007 | 1 mm x 10 well |
| | 16008 | 1 mm x 12 well |
| Polycarbonate Dual-sided Comb | 16013 | 1 mm x 8 well/16 well |
| | 16014 | 1 mm x 10 well/14 well |
| | 16015 | 2 mm x 8 well/16 well |
| | 16016 | 2 mm x 10 well/14 well |
| Teflon Single-sided Comb | 16009 16010 | 1 mm x 3 well 1 mm x 8 well |
| | 16010 | 1 mm x 8 well 1 mm x 10 well |
| | 16011 | 1 mm x 12 well |
| Teflon Dual-sided Comb | 16012 | 1 mm x 8 |
| | 16018 | well/16 well |
| | 10010 | well/14 well |
| | 16019 | 2 mm x 8 well/16 well |
| | 16020 | 2 mm x 10 well/14 well |
| Power Supply w/Timer, (120V) | 16023 | 1 unit |
| Power Supply w/Timer, (220V) | 16023-220 | 1 unit |
| 96 Well Plate Shaker (120V) | 11996 | 1 unit |
| 96 Well Plate Shaker (220V) | 11996-220 | 1 unit |
| Plate Adapter Set | 11999 | 1 set |
| Vacuum Pump (120V) | 11998 | 1 unit |
| Vacuum Pump (220V) | 11998-220 | 1 unit |
| UltraVac™ Manifold | 11997 | 1 unit |
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